

Tennessee Quality Milk Initiative

Bulk Tank Milk Quality

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Who is responsible for the safety and quality of dairy products? Dairy producers? Processors? Retailers? Consumers? In fact, all of these parties – from farm to fork – share in this responsibility. What starts as a high-quality product on the farm can be ruined somewhere along this chain. However, a poor-quality, inferior raw product leaving the farm cannot be transformed into a safe and high-quality product for the consumer. Despite technical advances in milk processing, the safety and quality of dairy products is still determined on the farm. Pasteurization does provide a certain level of safety, but is not the ultimate tool for protecting and satisfying consumers. Many aspects of milk safety and quality cannot be changed through pasteurization. The dairy industry continues to develop and adopt testing methods to gauge the safety, quality and suitability of raw milk for processing. It is vital that dairy producers understand these tests and their role in producing a safe, high-quality and nutritious product.

Dairy Product Safety

Bacteria are present in raw milk as a result of milking cows with mastitis, contamination from the environment during milking, contaminated milking and handling equipment and/or bacterial growth during storage. Pasteurization is an effective method of destroying mastitis-causing bacteria, foodborne pathogens and other bacteria in milk. However, it does not destroy **all** bacteria, and it does not destroy heat-stable toxins produced by some bacteria found in bulk tank milk. Several bulk tank milk studies have detected the presence of foodborne pathogens in bulk tanks (5, 6, 9, 11, 15, 19, 21, 22, 23, 25, 26, 28, 29, 30, 31, 32, 34, 40, 41, 42, 44). Outbreaks of disease in humans have been traced back to the consumption of raw **and** pasteurized milk (1, 2, 8, 10, 12, 13, 17). Some pathogens such as *Listeria monocytogenes* can survive and thrive for extended periods in reservoirs on processing equipment and are difficult to eradicate. This can lead to recontamination of dairy products (33, 45, 46). Though this may appear to be an issue only at the processing level, reducing foodborne pathogens in raw milk will ultimately reduce consumers' risk. Additionally, some strains of bacteria, specifically *Staphylococcus aureus*, produce toxins and have been reported to cause a number of disease or food poisoning outbreaks because of ingestion of contaminated milk or dairy products (1, 2, 13).

Dairy Product Quality

Bacteria in milk, whether originating from the cow or the environment, can significantly impact the quality of dairy products and therefore consumer acceptance. Many bacteria produce heat-stable enzymes. These enzymes are not affected by pasteurization and continue to cause damage to fat and/or protein in the final product. Enzymes that break down milk fat release free fatty acids, which can result in off-flavors and rancidity (3). Enzymes that break down milk protein can cause bitter flavors in milk. Because of bacterial enzyme activity before and after pasteurization, milk quality is altered, resulting in reduced shelf life (24).

Somatic Cell Count (SCC)

It has long been recognized that milk containing a high number of somatic cells will have reduced cheese yield. However, in a fluid milk market, such as in the South, there seems to be much debate over the value of SCC as a safety and quality parameter. At this time, there is no known direct human health concern with consuming milk containing a high number of somatic cells (white blood cells) from cows (16). However, relationships do exist between elevated SCC and other safety parameters, in particular antibiotic residues (36, 37, 38, 43). Thus, the SCC of milk is an *indirect* indicator of product safety. In sensory evaluations (i.e., taste-tests), pasteurized milk made from milk containing a high number of somatic cells (>500,000 cells/ml) scored lower than milk containing a lower SCC (250,000 cells/ml) (27) and also had a reduced shelf life (24). Cows with elevated SCCs have experienced an intramammary infection, likely caused by bacteria.

As stated earlier, enzymes produced by bacteria cause compositional changes in milk before and after pasteurization, which can affect quality. Therefore, SCCs are used more as an *indirect* indicator of quality. Some processors place a lot of emphasis on SCC limits. Although the Pasteurized Milk Ordinance (PMO) sets the legal limit at 750,000 cells/ml (2005), one processor in Tennessee has a quality standard of 350,000 cells/ml. At this time, other processors do not place as much emphasis on SCC but rather set quality standards based on bacteria counts.

Standard Plate Count (SPC)

The SPC is specified by the PMO as the official regulatory test used for estimating bacterial numbers in raw milk. For Grade A milk, the PMO requires a SPC < 100,000 colony forming units(cfu)/ml (2005). However, industry standards may be much lower than this. A SPC is run by incubating a sample at 89.6 degrees F (32 C) for 48 hours, followed by counting bacterial colonies (35). Since bacteria are miniscule in size, the incubation period provides bacteria an ideal environment to grow and proliferate to a size that can be detected and counted. However, there are problems associated with using the SPC alone. The SPC gives no indication as to the types of bacteria present and does not indicate the specific source of contamination. Moreover, the SPC may not give a complete count of all bacteria in milk, because some bacteria only grow at lower temperatures. So cold-loving bacteria are still present in the milk and could damage the product, but this would not be apparent with a SPC alone (7).

Preliminary Incubation Count (PIC)

The PIC has gained importance in quality testing as it measures bacteria that thrive in lower temperatures. Psychotrophic (i.e. cold-loving) bacteria can grow at temperatures from 32 to 68 degrees F (4) and are mostly comprised of gram-negative bacteria (pseudomonas, coliforms, flavobacterium and alcaligenes) (7).

The PIC is a two-step process. First, the sample is "pre-incubated" at 55 degrees F (12.7 C) for 18 hours. This allows cold-loving bacteria to grow and get a 'jump start' on bacteria that require warmer temperatures. The second step is to incubate at 89.6 degrees F (32 C) for 48 hours (a typical SPC) to allow the warm-loving bacteria to grow. Following incubation, all bacteria are counted (7). Because of this two-step approach, it is possible to conduct a SPC and a PIC on the same sample and get completely different results. However, this process also gives a more complete and accurate determination of the bacterial population in milk. Unfortu-

nately, as with the SPC, a PIC does not indicate the specific source of contamination and requires a very broad approach when problem-solving milk with high PICs.

Laboratory Pasteurized Count (LPC)

At one time, the LPC was an important quality parameter for many processors, but emphasis is now changing from LPCs to PICs. However, it is still important to recognize and understand this method of testing. The LPC is conducted by heating a sample to 145 degrees F (62.8 C) and holding it at that temperature for 30 minutes. This method simulates a low-temperature, long-time pasteurization process. After incubation, the number of bacteria are counted (35). The purpose of the LPC is to determine the levels of bacteria that can survive pasteurization. These heat-loving bacteria (thermoduric) are typically found in soil and often form spores, a survival mechanism making them resistant to many agents, including sanitizers (18). Thermoduric bacteria include: *Micrococcus*, *Microbacterium*, *Lactobacillus*, *Bacillus*, *Clostridium* and occasionally *Streptococci* (35). These bacteria retain their activity and can affect quality of a post-pasteurized product.

Coliform Count (CC)

The CC is performed by culturing dilutions of raw milk on selective media and incubating at 90 degrees F (32 C) for 24 hours to promote growth of coliform bacteria (35). Typically, the CC is used as an indicator of unsanitary production practices. High CCs can originate from dirty cows, poor milking hygiene, poor cleaning and/or sanitizing of equipment or bacterial growth on milking equipment. Additionally, CCs may also be influenced by coliform mastitis. This fact is somewhat dismissed, because typically, cows infected by coliforms either shed relatively low numbers or are acutely infected and their milk is not likely placed in the bulk tank (18, 20). However, in some coliform infections, the number of coliform bacteria can reach several million/ml in the infected quarter before clinical signs are visible (39). Milk from the infected cow would not be held out of the bulk tank until the next milking when signs of infection are apparent.

Conclusions

Providing a safe, high-quality and nutritious dairy product is challenging, because all aspects of the production chain, from the farm to the consumer must be considered. However, the initial responsibility rests with dairy producers that provide raw milk. A high incidence of mastitis (subclinical or clinical) and on-farm contamination of milk with bacteria found in the dairy farm environment leads to safety and quality issues during processing. The dairy industry has developed many direct and indirect measurements of raw milk safety and quality. A better understanding of these measurements will allow producers to adopt production practices that will result in a safer and higher-quality raw product that meets legal standards, industry-placed quality standards and consumer acceptance.

References

1. Adesiyun, A.A., L.A. Webb, and H.T. Romain. 1998. Prevalence and characteristics of *Staphylococcus aureus* strains isolated from bulk and composite milk and cattle handlers. *J. Food Prot.* 61:629.
2. Asao, T., Y. Kumeda, T. Kawai, T. Shibata, H. Oda, K. Haruki, H. Nakazawa, and S. Kozaki. 2003. An extensive outbreak of staphylococcal food poisoning due to low-fat milk in Japan: estimation of enterotoxin A in the incriminated milk and powdered skim milk. *Epidemiol. Infect.* 130:33.
3. Barbano, D.M., R.J. Verdi, A.I. Saeman, D.M. Galton, and R.R. Rasmussen. 1987. Impact of mastitis on dairy product yield and quality. 26th NMC Annual Meeting Proceedings. National Mastitis Council, Inc. Arlington, VA. p.132.

4. Cook, N.B. Localizing a SPC Problem. University of Wisconsin Extension Service. Available: <http://www.uwex.edu/milkquality/PDF/localspc.PDF>
5. Davidson, R.J., D.W. Sprung, C.E. Park, and M.K. Raymond. 1989. Occurrence of *Listeria monocytogenes*, *Campylobacter* spp., and *Yersinia enterocolitica* in Manitoba raw milk. *Can. Inst. Food Sci. Technol. J.* 22:70-74.
6. Doyle, M.P., and D.J. Roman. 1982. Prevalence and survival of *Campylobacter jejuni* in unpasteurized milk. *Appl. Environ. Microbiol.* 44:1154.
7. Ecolab. 2005. PI Test: What is it? Ecolab Inc., Food and Beverage Division. St. Paul, MN.
8. Evans, M.R., R.J. Roberts, C.D. Ribeiro, D. Gardner, and D. Kembrey. 1996. A milk-borne campylobacter outbreak following an educational farm visit. *Epidemiol. Infect.* 117:457.
9. Farber, J.M., G.W. Sander, and S.A. Malcolm. 1988. The presence of *Listeria* spp. in raw milk in Ontario. *Can. J. Microbiol.* 34:95-100.
10. Fahey, T., D. Morgan, C. Gunneburg, G.K. Adak, F. Majid, and E. Kaczmarek. 1995. An outbreak of *Campylobacter jejuni* enteritis associated with failed milk pasteurization. *J. Infect.* 31:137.
11. Fedio, W.M., and H. Jackson. 1990. Incidence of *Listeria monocytogenes* in raw milk in Alberta. *Can. Inst. Food Sci. Technol. J.* 23:236.
12. Fleming, D.W., S.L. Cochi, K.L. MacDonald, J. Brondum, P.S. Hayes, B.D. Plikaytis, M.B. Holmes, A. Audurier, C.V. Broome, and A.L. Reingold. 1985. Pasteurized milk as a vehicle of infection in an outbreak of listeriosis. *N. Engl. J. Med.* 312:404.
13. Genigeorgis, C.A. 1989. Present state of knowledge on staphylococcal intoxication. *Int. J. Microbiol.* 9:327.
14. Grade "A" Pasteurized Milk Ordinance, 2005 Revision. U.S. Department of Health & Human Services, Public Health Service, Food and Drug Administration.
15. Hassan, L., H.O. Mohammed, P.L. McDonough, and R.N. Gonzalez. 2000. A cross-sectional study on the prevalence of *Listeria monocytogenes* and *Salmonella* in New York dairy herds. *J. Dairy Sci.* 83:2441.
16. Hogan, J. 2005. Human health risks associated with high SCC milk: Symposium overview. 44th NMC Annual Meeting Proceedings. National Mastitis Council, Inc. Verona, WI. pp. 73-75.
17. Hutchinson, D.N., F.J. Bolton, P.M. Hinchliffe, H.C. Dawkins, S.D. Horsley, E.G. Jessop, P.A. Robertshaw, and D.E. Counter. 1985. Evidence of udder excretion of *Campylobacter jejuni* as the cause of milk-borne campylobacter outbreak. *J. Hyg. (Lond.)* 94:205.
18. Ingall, W. 1998. Milk quality and factors influencing the production of high quality milk. West Agro, Inc. Kansas City, MO.
19. Jayarao, B.M., and D.R. Henning. 2001. Prevalence of foodborne pathogens in bulk tank milk. *J. Dairy Sci.* 84:2157.
20. Kirk, J.H. 2005. Milk quality on the dairy – who is responsible? University of California Cooperative Extension. Available: <http://www.vetmed.ucdavis.edu/vetext/INF-DA.HTML#MQ>
21. Liewen, M.B., and M.W. Plautz. 1988. Occurrence of *Listeria monocytogenes* in raw milk in Nebraska. *J. Food Prot.* 51:840.
22. Lovett, J., D.W. Francis, and J.M. Hunt. 1983. Isolation of *Campylobacter jejuni* from raw milk. *Appl. Environ. Microbiol.* 46:459.
23. Lovett, J., D.W. Francis, and J.M. Hunt. 1987. *Listeria monocytogenes* in raw milk: detection, incidence, and pathogenicity. *J. Food Prot.* 50:188.
24. Ma, Y., C. Ryan, D.M. Barbano, D.M. Galton, M.A. Rudan, and K.J. Boor. 2000. Effects of somatic cell count on quality and shelf-life of pasteurized fluid milk. *J. Dairy Sci.* 83:264-274.
25. McEwen, S.A., S.W. Martin, R.C. Clarke, S.E. Tamblyn, and J.J. McDermott. 1988. The prevalence, incidence, geographical distribution, and antimicrobial sensitivity patterns and plasmid profiles of milk filter *Salmonella* isolates from Ontario dairy farms. *Can. J. Vet. Res.* 52:18.
26. McManus, C., and J.M. Lanier. 1987. *Salmonella*, *Campylobacter jejuni*, and *Yersinia enterocolitica* in raw milk. *J. Food Prot.* 50:51.
27. Mitchell, G.E., Rogers, S.A., and S.L. Slattery. 1987. Research on the effects of somatic cell count on milk composition and milk product quality. *Dairy Products.* 14:15-16.
28. Murinda, S.E., L.T. Nguyen, S.J. Ivey, B.E. Gillespie, R.A. Almeida, F.A. Draughon, and S.P. Oliver. 2002a. Molecular characterization of *Salmonella* spp. isolated from bulk tank milk and cull dairy cow fecal samples. *J. Food Prot.* 65:1100.
29. Murinda, S.E., L.T. Nguyen, S.J. Ivey, B.E. Gillespie, R.A. Almeida, F.A. Draughon, and S.P. Oliver. 2002b. Prevalence and molecular characterization of *Escherichia coli* O157:H7 in bulk tank milk and fecal samples from cull cows: a 12-month survey of dairy farms in east Tennessee. *J. Food Prot.* 65:752.
30. Murinda, S.E., L.T. Nguyen, T.L. Landers, F.A. Draughon, A.G. Mathew, J.S. Hogan, K.L. Smith, D.D. Hancock, and S.P. Oliver. 2004a. Comparison of *Escherichia coli* isolates from humans, food, farm and companion animals for presence of Shiga-toxin producing *Escherichia coli* virulence markers. *Foodborne Pathogens & Disease.* 1:178.
31. Murinda, S.E., L.T. Nguyen, H.M. Nam, R.A. Almeida, S.J. Headrick, and S.P. Oliver. 2004b. Detection of sorbitol-negative and sorbitol-positive Shiga-toxin producing *Escherichia coli*, *Listeria monocytogenes*, *Campylobacter jejuni* and *Salmonella* spp. in dairy environmental samples. *Foodborne Pathogens & Disease.* 1:97.
32. Oliver, S.P., B.M. Jayarao, and R.A. Almeida. 2005. Foodborne pathogens in milk and the dairy farm environment: Food safety and public health implications. *Foodborne Pathogens and Disease.* p. 115-129.
33. Pritchard, T.J., K.J. Flanders, and C.W. Donnelly. 1995. Comparison of the incidence of *Listeria* on equipment versus environmental sites within dairy processing plants. *Int. J. Food Microbiol.* 26:375.
34. Rohrbach, B.W., F.A. Draughon, P.M. Davidson, and S.P. Oliver. 1992. Prevalence of *Listeria monocytogenes*, *Campylobacter jejuni*, *Yersinia enterocolitica*, and *Salmonella* in bulk tank milk: risk factors and risk of human exposure. *J. Food Prot.* 55:93.
35. Ruegg, P.L., and D.J. Reinemann. 2002. Milk quality and mastitis tests. University of Wisconsin Extension Service.
36. Ruegg, P.L., and T.J. Tabone. 2000. The relationship between antibiotic residue violations and somatic cell counts in Wisconsin dairy herds. *J. Dairy Sci.* 83:2805-2809.
37. Sargeant, J.M., Y.H. Schukken, and K.E. Leslie. 1998. Ontario bulk milk somatic cell count reduction program: progress and outlook. *J. Dairy Sci.* 81:1545-1554.
38. Saville, W.J.A., T.E. Wittum, and K.L. Smith. 2000. Association between measures of milk quality and risk of violative antimicrobial residues in grade-A milk. *J. Am. Vet. Med. Assoc.* 217:541-545.
39. Schroeder, J.W. 1997. Mastitis control program: Milk quality evaluation tools for dairy farmers. North Dakota State University Extension Service. AS-1131.
40. Slade, P.J., D.L. Collins-Thompson, and F. Fletcher. 1988. Incidence of *Listeria* species in Ontario raw milk. *Can. Inst. Food Sci. Technol. J.* 21:425.

41. Steele, M.L., W.B. McNab, C. Poppe, M.W. Griffiths, S. Chen, S.A. Degrandis, L.C. Fruhner, C.A. Larkin, J.A. Lynch, and J.A. Odermeru. 1997. Survey of Ontario bulk tank milk for foodborne pathogens. *J. Food Prot.* 60:1341.
42. Van Kessel, J.A., J.S. Karns, L. Gorski, B.J. McCluskey, and M.L. Perdue. 2004. Prevalence of *Salmonellae*, *Listeria monocytogenes* and fecal coliforms in bulk tank milk on U.S. dairies. *J. Dairy Sci.* 87:2822-2830.
43. Van Schaik, G., M. Lotem, and Y.H. Schukken. 2002. Trends in somatic cell counts, bacterial counts, and antibiotic residue violations in New York State during 1999-2000. *J. Dairy Sci.* 85:782-798.
44. Waak, E., W. Tham, and M.L. Danielsson-Tham. 2002. Prevalence of *Listeria monocytogenes* strains isolated from raw whole milk in farm bulk tanks and dairy plants receiving tanks. *Appl. Environ. Microbiol.* 68:3366.
45. Walker, R.L., L.H. Jensen, H. Kinde, A.V. Alexander, and L.S. Owen. 1991. Environmental survey for *Listeria* species in a frozen milk products plant in California. *J. Food Prot.* 54:178.
46. Wong, A.C. 1998. Biofilms in food processing environments. *J. Dairy Sci.* 81:2765.